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NEWS 12 FEB 02 GENBANK enhanced with SET PLURALS and SET SPELLING  
  
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AND CURRENT DISCOVER FILE IS DATED 23 JUNE 2008.  
  
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AGGREGATION)

=> s (acid or pH 5 or pH 6) (4A) (precipitation or flocculation or aggregation)

L2 22202 (ACID OR PH 5 OR PH 6) (4A) (PRECIPITATION OR FLOCCULATION OR  
AGGREGATION)

=> s (protein or enzyme or polypeptide) (4A) (preparation or isolation or  
purification or separation or production)

L3 232431 (PROTEIN OR ENZYME OR POLYPEPTIDE) (4A) (PREPARATION OR ISOLATIO  
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=> s l1 and l2 and l3

L4 10 L1 AND L2 AND L3

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L5 ANSWER 1 OF 8 EMBASE COPYRIGHT (c) 2009 Elsevier B.V. All rights  
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AN 2005232553 EMBASE

TI Characterization of the thermophilic isoamylase from the thermophilic  
archaeon Sulfolobus solfataricus ATCC 35092.

AU Fang, Tsuei-Yun (correspondence); Yu, Ching-Ju; Shih, Tong-Yuan

CS Department of Food Science, National Taiwan Ocean University, 2 Pei-Ning  
Road, Keelung 202, Taiwan, Province of China. tyfang@mail.ntou.edu.tw

AU Tseng, Wen-Chi

CS Department of Chemical Engineering, National Taiwan University of Science  
and Technology, Taipei, Taiwan, Province of China.

SO Journal of Molecular Catalysis B: Enzymatic, (1 Jun 2005) Vol. 33, No.  
3-6, pp. 99-107.

Refs: 40

ISSN: 1381-1177 CODEN: JMCEF8

PUI S 1381-1177(05)00044-5

CY Netherlands

DT Journal; Article  
 FS 029 Clinical and Experimental Biochemistry  
 LA English  
 SL English  
 ED Entered STN: 16 Jun 2005  
 Last Updated on STN: 16 Jun 2005  
 AB Isoamylase catalyzes the hydrolysis of  $\alpha$ -1,6-glucosidic linkages of starch and related polysaccharides. In this study, the treX gene (GenBank accession number AE006815 REGION: 9279 ... 11435) encoding the thermophilic isoamylase was PCR-cloned from the genomic DNA of Sulfolobus solfataricus ATCC 35092 to an expression vector with a T7lac promoter. Both wild-type and His-tagged isoamylases were expressed in Escherichia coli. The wild-type isoamylase was purified sequentially using heat treatment, nucleic acid precipitation, ion-exchange chromatography, and gel filtration chromatography while the His-tagged isoamylase was purified from the cell-free extract directly by metal chelating chromatography. Both enzymes were active only under their homo-trimer forms. In the absence of NaCl, both enzymes became inactive monomers. In addition, both enzymes were more stable when being stored at room temperature than at 4°C. They had an apparent optimal pH of 5 and an optimal temperature at 75°C. The enzyme activities remained unchanged after a 2 h incubation at 80 and 75°C for the wild-type and His-tagged enzymes, respectively. These thermophilic isoamylases showed a potential to be used in industry to degrade the branching points of starch at a high temperature. .COPYRG. 2005 Elsevier B.V. All rights reserved.

L5 ANSWER 2 OF 8 CAPLUS COPYRIGHT 2009 ACS on STN  
 AN 2004:965380 CAPLUS  
 DN 141:406798  
 TI Lactobacillus acidophilus nucleic acid sequences encoding cell surface protein homologs and the therapeutic and food uses  
 IN Klaenhammer, Todd R.; Russell, William M.; Alterman, Eric; Cano, Raul J.; Hamrick, Alice  
 PA North Carolina State University, USA  
 SO PCT Int. Appl., 722 pp.  
 CODEN: PIXXD2  
 DT Patent  
 LA English  
 FAN.CNT 1

	PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
PI	WO 2004096992	A2	20041111	WO 2004-US12717	20040423
	WO 2004096992	A3	20060216		
	W:				
	AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW				
	RW:				
	BW, GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PL, PT, RO, SE, SI, SK, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG				
	AU 2004235346	A1	20041111	AU 2004-235346	20040423
	US 20050112612	A1	20050526	US 2004-831070	20040423
	US 7348420	B2	20080325		
	EP 1658303	A2	20060524	EP 2004-750608	20040423
	R:				
	AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR, BG, CZ, EE, HU, PL, SK, HR				

	JP 2006525805	T	20061116	JP 2006-513298	20040423
	US 20070172495	A1	20070726	US 2007-701319	20070201
	US 20070258955	A1	20071108	US 2007-701335	20070201
PRAI	US 2003-465621P	P	20030425		
	US 2004-831070	A3	20040423		
	WO 2004-US12717	W	20040423		

AB Cell wall, cell surface, and secreted protein nucleic acid mols. and polypeptides and fragments and variants thereof are disclosed in the current invention. In addition, cell wall, cell surface and secreted fusion proteins, antigenic peptides, and anti-cell wall, cell surface and secreted antibodies are encompassed. The invention also provides recombinant expression vectors containing a nucleic acid mol. of the invention and host cells into which the expression vectors have been introduced. Methods for producing the polypeptides of the invention and methods for their use are further disclosed. Their isolation and characterization will aid in developing essential probiotic products with numerous applications, including those that benefit human or animal health, and those concerned with food production and safety.

L5 ANSWER 3 OF 8 EMBASE COPYRIGHT (c) 2009 Elsevier B.V. All rights reserved on STN

AN 2001278595 EMBASE

TI Proteins from fish eggs that protect DNA from acid precipitation and inhibit DNA synthesis.

AU Tsamis, Vaggelis; Mamuris, Zissis; Panagiotaki, Panagiota; Kouretas, Demetrios (correspondence)

CS School of Agriculture, University of Thessaly, Fitoko Neas Ionias, Magnisia 38446, Greece. dkouret@uth.gr

SO Comparative Biochemistry and Physiology - C Toxicology and Pharmacology, (2001) Vol. 129, No. 4, pp. 369-376.

Refs: 10

ISSN: 1532-0456 CODEN: CBPPFK

PUI S 1532-0456(01)00215-0

CY United States

DT Journal; Article

FS 029 Clinical and Experimental Biochemistry

LA English

SL English

ED Entered STN: 23 Aug 2001

Last Updated on STN: 23 Aug 2001

AB We partially characterized proteins that inhibit DNA acid precipitation from various fish eggs (*Sparus aurata*, *Dicentrarchus labrax*, *Mugil cephalus* and *Zeus faber*). The active proteins were purified by acetone fractionation. The activity was found to be heat resistant. Of bivalent cations tested only Co(2+) and Cu(2+) exerted a profound promoting effect in the activity from all fish. The protein fraction from *Sparus aurata* inhibited DNA synthesis in PCR performed by different DNA polymerases. The possible role of DNA protective proteins in fish egg physiology is discussed. .COPYRG. 2001 Elsevier Science Inc. All rights reserved.

L5 ANSWER 4 OF 8 EMBASE COPYRIGHT (c) 2009 Elsevier B.V. All rights reserved on STN DUPLICATE 1

AN 2000081406 EMBASE

TI Microbial production, purification, and characterization of (S)-specific N-acetyl-2-amino-1-phenyl-4-pentene amidohydrolase from *Rhodococcus globerulus* K1/1.

AU Wahl, P.; Walser-Volken, P.; Laumen, K.; Kittelmann, M. (correspondence); Ghisalba, O.

CS Novartis Pharma AG, Research Department, Core Technology Area, WSJ-508.102 A, CH-4002 Basel, Switzerland. matthias.kittelmann@pharma.novartis.com

AU Kittelmann, M. (correspondence)  
 CS Novartis Pharma AG, Research Department, Core Technology Area, CH-4002  
 Basel, Switzerland. matthias.kittelmann@pharma.novartis.com  
 SO Applied Microbiology and Biotechnology, (1999) Vol. 53, No. 1, pp. 12-18.  
 Refs: 22  
 ISSN: 0175-7598 CODEN: AMBIDG  
 CY Germany  
 DT Journal; Article  
 FS 004 Microbiology: Bacteriology, Mycology, Parasitology and Virology  
 LA English  
 SL English  
 ED Entered STN: 16 Mar 2000  
 Last Updated on STN: 16 Mar 2000  
 AB Rhodococcus globerulus K1/1 was found to express an inducible (S)-  
 specific N-acetyl-2-amino-1-phenyl-4-pentene amidohydrolase. Optimal  
 bacterial growth and amidohydrolase expression were both observed at about  
 pH 6.5. Purification of the enzyme to a single band  
 in a Coomassie blue-stained SDS-PAGE gel was achieved by nucleic  
 acid and ammonium sulfate precipitation of Rhodococcus  
 globerulus K1/1 crude extract and column chromatography on TSK  
 Butyl-650(S) Fractogel and Superose 12HR. The amidohydrolase was purified  
 to a homogeneity leading to a tenfold increase of the specific activity  
 with a recovery rate of 65%. At pH 7.0 and 23 °C the enzyme showed  
 no loss of activity after 30 days incubation. The amidohydrolase was  
 stable up to 55 °C. The enzyme was inhibited strongly only by 10  
 mM Zn(2+) among the tested metal cations and was inhibited 100% by 0.01 mM  
 phenylmethanesulfonyl fluoride. The molecular weight of the native enzyme  
 was estimated to be 92 kDa by gel filtration and 55 kDa by SDS-PAGE,  
 suggesting a homodimeric structure.

L5 ANSWER 5 OF 8 MEDLINE on STN DUPLICATE 2  
 AN 1984193740 MEDLINE  
 DN PubMed ID: 6371782  
 TI [Purification and properties of phenylalanyl-tRNA-synthetase from  
 Escherichia coli MRE-600].  
 Ochistka i nekotorye svoistva fenilalanil-tRNK-sintetazy iz Escherichia  
 coli MRE-600.  
 AU Ankilova V N; Lavrik O I; Khodyreva S N  
 SO Prikladnaia biokhimiia i mikrobiologiya, (1984 Mar-Apr) Vol. 20, No. 2,  
 pp. 208-16.  
 Journal code: 0023416. ISSN: 0555-1099.  
 CY USSR  
 DT (ENGLISH ABSTRACT)  
 Journal; Article; (JOURNAL ARTICLE)  
 LA Russian  
 FS Priority Journals  
 EM 198406  
 ED Entered STN: 19 Mar 1990  
 Last Updated on STN: 6 Feb 1998  
 Entered Medline: 14 Jun 1984  
 AB A preparative scale method for isolation of highly purified  
 phenylalanyl-tRNA synthetase from E. coli MRE-600 was developed. It  
 consists of cell destroying, nucleic acid  
 precipitation with streptomycin sulfate, fractionation with  
 ammonium sulfate followed by chromatography on different carriers  
 (Sephadex G-200, DEAE-cellulose, DEAE-Sephadex A-50, and hydroxyapatite).  
 The mode of cell destroying was found to affect the process of the further  
 enzyme purification. The phenylalanyl-tRNA synthetase  
 was purified 540-fold, with recovery being 20.6% and the specific activity  
 - 540 units per mg protein. The enzyme content in the purified  
 preparation was 80-90% judging by electrophoresis in PAAG. The

molecular weights of the subunits determined by electrophoresis under denaturative conditions were found to be 102,000 +/- 4000 (beta) and 42,000 +/- 2000 (alpha). The molecular weight of the native enzyme determined by gel filtration through Sephadex G-200 and electrophoresis at varied concentrations of polyacrylamide was found to be 340,000 +/- 20,000. The Km values for tRNA, ATP and phenylalanine in the aminoacylation reaction are equal to  $5.4 \times 10^{-7}$  M,  $1.9 \times 10^{-4}$  M, and  $3.7 \times 10^{-6}$  M, respectively.

L5 ANSWER 6 OF 8 BIOSIS COPYRIGHT (c) 2009 The Thomson Corporation on STN  
AN 1979:264554 BIOSIS  
DN PREV197968067058; BA68:67058  
TI NOVEL METHOD FOR THE REDUCTION OF NUCLEIC-ACID IN YEAST PROTEIN.  
AU SHETTY K J [Reprint author]; KINSELLA J E  
CS DEP FOOD SCI, COLL AGRIC LIFE SCI, CORNELL UNIV, ITHACA, NY 14853, USA  
SO Biotechnology and Bioengineering, (1979) Vol. 21, No. 2, pp. 329-332.  
CODEN: BIBIAU. ISSN: 0006-3592.  
DT Article  
FS BA  
LA ENGLISH  
AB The succinylation of the protein increased with increasing concentrations of succinic anhydride and more than 90% modification was obtained at a 1:1 ratio of succinic anhydride to protein. The nucleic acid content of the precipitated protein progressively decreased with the extent of succinylation. The data were consistent with the suggestion that succinylation of the protein, and perhaps the nucleic acid, accentuated the differences in solubility between the protein and RNA over a narrow pH range. This was most pronounced between pH 4-4.5 where over 95% of the nucleic acid remained in solution during precipitation of the modified protein. Maximum cell protein recovery (i.e., 80%) was obtained at pH 4.2-4.3, which was greater than that recovered from control cells [*Saccharomyces carlsbergensis*], i.e., those not treated with succinic anhydride. The nucleic acid content of the succinylated protein recovered between pH 4.2-4.4, averaged 1.8% on a dry-weight basis. The protein isolate (92% protein) recovered from yeast cells by this process is highly soluble and possesses many of the desirable physicochemical properties required in food proteins. This method is very amenable to methods currently in use for the isolation of protein from yeast cells and represents a significant development that should facilitate the exploitation of yeast as a source of food-grade protein.

L5 ANSWER 7 OF 8 BIOSIS COPYRIGHT (c) 2009 The Thomson Corporation on STN  
AN 1978:195580 BIOSIS  
DN PREV197866008077; BA66:8077  
TI INVESTIGATION OF THE UNIT OPERATIONS INVOLVED IN THE CONTINUOUS FLOW ISOLATION OF BETA GALACTOSIDASE FROM ESCHERICHIA-COLI.  
AU HIGGINS J J [Reprint author]; LEWIS D J; DALY W H; MOSQUEIRA F G; DUNNILL P; LILLY M D  
CS BIOSCI LAB, SHELL RES LTD, SITTINGBOURNE, KENT, ENGL, UK  
SO Biotechnology and Bioengineering, (1978) Vol. 20, No. 2, pp. 159-182.  
CODEN: BIBIAU. ISSN: 0006-3592.  
DT Article  
FS BA  
LA ENGLISH  
AB A 1000 l fermentor was used to produce a continuous feed of *E. coli* containing a high level of  $\beta$ -galactosidase. The individual unit operations was investigated for the isolation of the enzyme: cell disruption, nucleic acid removal, protein precipitation and solid-liquid separation after each stage. The information obtained enabled the

operation of a semicontinuous process which when fully continuous would yield 100 g protein/h, comprising 23%  $\beta$ -galactosidase.

L5 ANSWER 8 OF 8 BIOSIS COPYRIGHT (c) 2009 The Thomson Corporation on STN  
AN 1955:23141 BIOSIS  
DN PREV19552900023197; BA29:23197  
TI The preparation of ribonucleo-protein from yeast.  
AU BONAR, ROBERT A.; DUGGAN, EDWARD L.  
CS U. California Sch. Med., Berkeley  
SO JOUR BIOL CHEM, (1955) Vol. 212, No. 2, pp. 697-704.  
DT Article  
FS BA  
LA Unavailable  
ED Entered STN: May 2007  
Last Updated on STN: May 2007  
AB Bakers' yeast (*Saccharomyces cerevisiae*) was ground with levigated alumina and extracted with water. The ground yeast suspension was separated centrifugally into 4 fractions, which were subjected to analysis for ribonucleic acid (RNA) N and P. The RNA assay consisted of perchloric acid extraction, alkaline hydrolysis, acidic precipitation of deoxypentose nucleic acid (DNA) and estimation of the total soluble nucleotide (RNA) using comparative spectrophotometry (260 m). Thymus DNA was found to yield appreciable derived fragments absorbing at this wavelength, unless temperature and time of contact with acid were closely controlled. The RNA was concentrated in 2 fractions, the submicroscopic particles and the coarse debris. Intermediate size particles and the supernatant fluid were relatively poor in RNA. A ribonucleoprotein material was isolated by precipitation at pH 4 and solution at pH 7. Its composition resembled that of the submicroscopic particle fraction. In connection with centrifugal fractionation with various rotors and centrifuges, the authors favor the use of the Performance Index (Pi), as defined by E. G. Pickets (Manual, Specialized Instrument Company, Belmont, Calif.). More precise comparison between centrifuges and heads is possible, since both maximal radius (R2) and minimal radius (R1) are included in the relation: [image].  
ABSTRACT AUTHORS: Authors

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